

Influenza B Viruses Hemagglutinin (HA) Specific ELISA Kit

Pack Size: 96 tests

Catalog Number: RAS-A207

IMPORTANT: Please carefully read this manual before performing your experiment.

For Research Use Only. Not For Use in Diagnostic or Therapeutic Procedures

INTENDED USE

This kit is developed for specific quantitative detection of influenza B virus HA protein in samples. This kit has been tested to specifically identify influenza B virus Victoria and Yamagata lineages HA protein. It is intended for research use only (RUO).

PRINCIPLE OF THE ASSAY

Influenza, commonly known as 'the flu', is an infectious disease of birds and mammals caused by RNA viruses of the family Orthomyxoviridae, the influenza viruses. The virus is divided into three main types (Influenzavirus A, Influenzavirus B, and Influenzavirus C), which are distinguished by differences in two major internal proteins (hemagglutinin (HA) and neuraminidase (NA), which are the most important targets for the immune system. Hemagglutinin binds to the sialic acid-containing receptors on the surface of host cells during initial infection and at the end of an infectious cycle which makes it a great target for vaccine studies.

This assay kit is used to measure the levels of influenza B virus HA protein by employing a standard sandwich-ELISA format. The microplate in the kit has been pre-coated with Anti-Influenza B (HA) Antibody. First add the standard samples provided in the kit and your samples to the plate, incubate and wash the wells. Then add the HRP-Anti-Influenza B (HA) Antibody to the plate, incubate and wash the wells. Lastly load the substrate into the wells and monitor color development in proportion with the amount of influenza B virus HA protein present. The reaction is stopped by the addition of a stop solution and the intensity of the absorbance can be measured at 450 nm and 630 nm. The OD Value reflects the amount of influenza B virus HA protein bound.

MATERIALS PROVIDED

TABLE 1. MATERIALS PROVIDED

| Catalog | Components | Size (96 tests) | Format | Storage | |
|------------|--|-----------------|--------|--------------------|--------------------|
| | | | | Unopened | Opened |
| RAS207-C01 | Pre-coated Anti-Influenza B (HA) Antibody Microplate | 1 plate | Solid | 2-8°C | 2-8°C |
| RAS207-C02 | Influenza B/Victoria Lineage (HA) Standard | 20 µg | Powder | 2-8°C | -70°C |
| RAS207-C03 | HRP-Anti-Influenza B (HA) Antibody | 20 µg | Powder | 2-8°C, avoid light | -70°C, avoid light |

| | | | | | |
|------------|--------------------|-------|--------|-----------------------|-----------------------|
| RAS207-C04 | 10×Washing Buffer | 50 mL | Liquid | 2-8°C | 2-8°C |
| RAS207-C05 | 2×Dilution Buffer | 50 mL | Liquid | 2-8°C | 2-8°C |
| RAS207-C06 | Substrate Solution | 12 mL | Liquid | 2-8°C, avoid light | 2-8°C, avoid light |
| RAS207-C07 | Stop Solution | 7 mL | Liquid | 2-8°C | 2-8°C |

REAGENTS/EQUIPMENT NEEDED BUT NOT SUPPLIED

Single or dual wavelength microplate reader with 450 nm and 630 nm filter;

Centrifuge;

37°C Incubator;

10 µL, 200 µL and 1000 µL precision pipettes;

10 µL, 200 µL and 1000 µL pipette tips;

Multichannel pipettes;

Tubes;

Graduated cylinder to prepare Wash Solution;

Deionized or distilled water to dilute 10×Washing Buffer;

STORAGE

1. Unopened kit should be stored at 2°C-8°C upon receiving.
2. Find the expiration date on the outside packaging and do not use reagents past their expiration date.
3. The opened kit should be stored per components table. The shelf life is 30 days from the date of opening.

REAGENT PREPARATION

1. Bring all reagents and samples to room temperature (20°C-25°C) before use. If crystals have formed in buffer solution, place the sample in a 37 °C incubator until the crystals have completely dissolved and bring the solution back to room temperature before use.
2. Reconstitute the provided lyophilized materials to stock solutions with distilled, sterile water as recommended in Table 2 and place the materials for 15 to 30 minutes at room temperature with occasional gentle mixing. Avoid vigorous shaking. The reconstituted stock solutions should be stored at -70°C. It is recommended not to freeze-thaw

more than 1 time, the packing specification shall not be less than 5 µg.

TABLE 2. RECONSTITUTION METHODS FOR 96 TESTS

| ID | Components | Size | Stock Solution Con. | Reconstitution Buffer and |
|------------|--|-------|---------------------|---------------------------|
| RAS207-C02 | Influenza B/Victoria Lineage (HA) Standard | 20 µg | 100 µg/mL | 200 µL water |
| RAS207-C03 | HRP-Anti-Influenza B (HA) Antibody | 20 µg | 100 µg/mL | 200 µL water |

RECOMMENDED SAMPLE PREPARATION

1. Working Fluid Preparation

1.1 Preparation of 1×Washing Buffer:

Dilute 50 mL 10×Washing Buffer with ultrapure water/deionized water to 500 mL.

1.2 Preparation of 1×Dilution Buffer:

Dilute 50 mL 2×Dilution Buffer with 1×Washing Buffer to 100 mL.

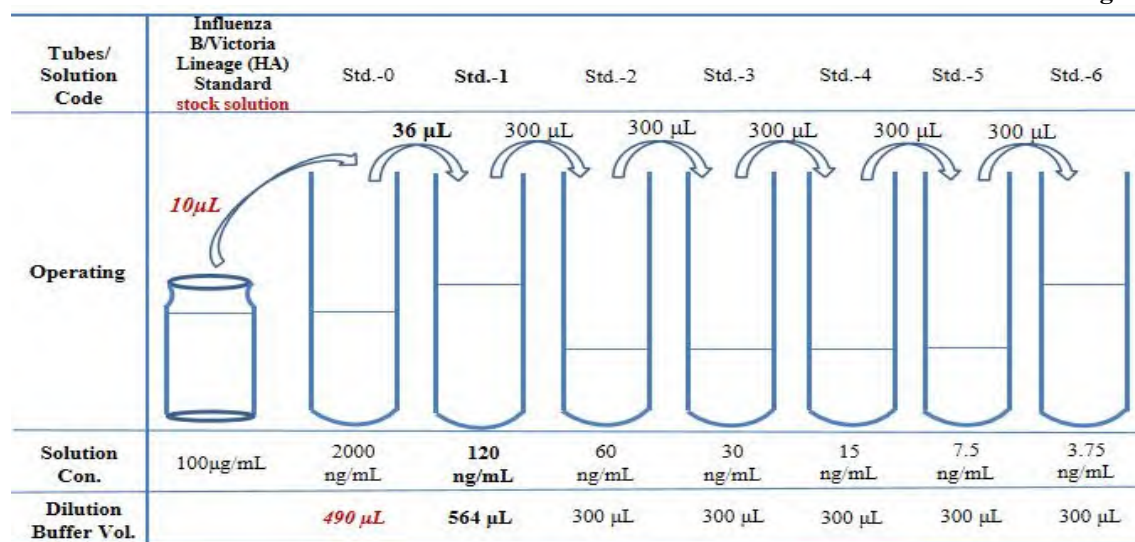
1.3 Preparation of HRP-Anti-Influenza B (HA) Antibody working fluid:

Dilute HRP-Anti-Influenza B (HA) Antibody to 1.0 µg/mL with Dilution Buffer. The prepared working fluid should avoid light. Please prepare it for one-time use only.

2. Preparation of Standard Curve

Make serial dilutions of the Influenza B/Victoria Lineage (HA) as a Standard curve with Dilution Buffer as recommended in Figure 1.

FIGURE 1. PREPARATION OF 1:1 SERIAL DILUTIONS OF THE Influenza B/Victoria Lineage (HA)



3. Add Samples

Add 100µL serially diluted **Influenza B/Victoria Lineage (HA)** Standard curve and samples to each well. For blank Control wells, please add 100µL 1×Dilution Buffer. Seal the plate with microplate sealing film and incubate at room temperature for 1.0 hour.

Note: It is recommended to set double holes for samples and standard curves to be tested.

4. Washing

Remove the remaining solution by aspiration, add 300 µL of 1×Washing Buffer to each well, soak for 30s, remove any remaining 1×Washing Buffer: by aspirating or decanting, invert the plate and blot it against paper towels. Repeat the wash step above for three times.

5. Add HRP-Anti-Influenza B (HA) Antibody

For all wells, add 100 µL **HRP-Anti-Influenza B (HA) Antibody (dilute to 1.0 µg/mL)** working solution. Seal the plate with microplate sealing film and incubate at room temperature for 1.0 hour.

6. Washing

Repeat step 4.

7. Substrate Reaction

Add 100 µL **Substrate Solution** to each well. Seal the plate with microplate sealing film and incubate at room temperature for 20 min, avoid light.

8. Termination

Add 50 µL **Stop Solution** to each well and tap the plate gently to allow thorough mixing.

Note: the color in the wells should change from blue to yellow.

9. Data Recording

Read the absorbance at 450 nm and 630 nm using UV/Vis microplate spectrophotometer within 5 minutes.

Note: To reduce the background noise, subtract the value read at $OD_{450\text{ nm}}$ with the value read at $OD_{630\text{ nm}}$.

CALCULATION OF RESULTS

1. Normal range of Standard curve: $R^2 \geq 0.9900$, detection range: 3.75-120 ng/mL.
2. If the OD value of the sample to be tested is higher than the highest standard, the sample shall be diluted with

dilution buffer and assay repeated.

3. To calibrate absorbance value obtained by the standard curve, the OD value of the sample to be measured is subtracted from the OD value of the blank control. The standard curve is plotted with the standard concentration as x-axis and the calibrated absorbance value as y-axis. Four parameters logistic are used to draw the standard curve and calculate the sample concentration.

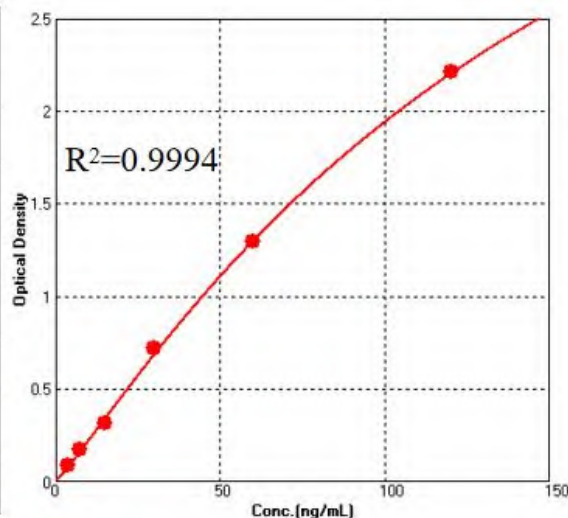
PRECAUTIONS

1. This kit is for research use only and is not for use in diagnostic or therapeutic procedures.
2. The kit should be used according to the instructions.
3. Do not mix reagents from different lots.
4. Bring all reagents and samples to room temperature (20 °C -25 °C) before use. If crystals have formed in buffer solution, warm to room temperature until the crystals have completely dissolved.
5. The kit should be stored at 2°C to 8°C.

TYPICAL DATA

The following data is for reference only. The sample concentration was calculated based on the results of the standard curve.

| Standard (ng/mL) | O.D.-1 | O.D.-2 | Average | Corrected |
|------------------|--------|--------|---------|-----------|
| 120 | 2.248 | 2.252 | 2.250 | 2.209 |
| 60 | 1.260 | 1.409 | 1.335 | 1.293 |
| 30 | 0.765 | 0.765 | 0.765 | 0.724 |
| 15 | 0.340 | 0.370 | 0.355 | 0.314 |
| 7.5 | 0.211 | 0.218 | 0.215 | 0.173 |
| 3.75 | 0.130 | 0.131 | 0.131 | 0.089 |
| 0 | 0.038 | 0.045 | 0.042 | / |



PRECISION

1. Intra-assay Precision

Three samples of known concentration were tested ten times on one plate to assess intra-assay precision.

2. Inter-assay Precision

Three samples of known concentration were tested in three separate assays to assess inter-assay precision.

| Sample | Intra-assay Precision | | | Inter-assay Precision | | |
|--------------|-----------------------|--------|-------|-----------------------|--------|-------|
| | 1 | 2 | 3 | 1 | 2 | 3 |
| n | 10 | 10 | 10 | 3 | 3 | 3 |
| Mean (ng/mL) | 88.792 | 43.058 | 8.834 | 91.972 | 44.366 | 8.966 |
| SD | 2.966 | 1.169 | 0.190 | 4.836 | 2.371 | 0.525 |
| CV (%) | 3.3 | 2.7 | 2.2 | 5.3 | 5.3 | 5.9 |

Note: The example data is for reference only.

RECOVERY

Three samples with different concentrations were tested to calculate the recovery rate.

| Sample(n=5) | Average Recovery % | Range % |
|-------------|--------------------|------------|
| High | 93.8 | 87.6-100.7 |
| Middle | 97.3 | 88.3-108.7 |
| Low | 95.2 | 84.3-107.2 |

LINEARITY

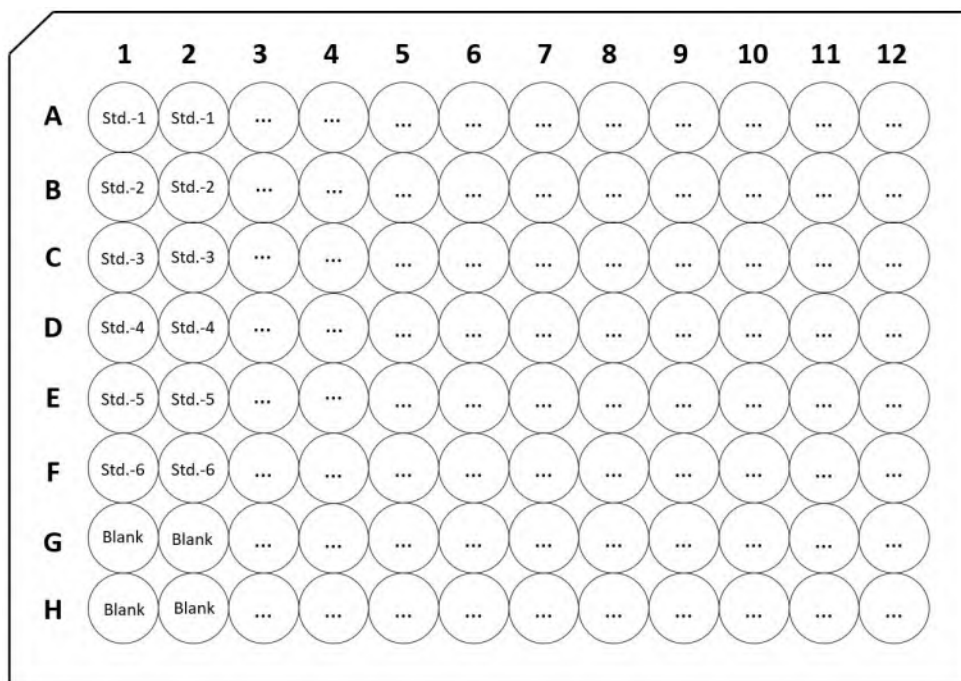
To assess the linearity of the assay, samples spiked with high concentrations were serially diluted with calibrator diluent to produce samples with values within the dynamic range of the assay.

| | | Cell culture medium (DMEM) | Cell culture medium (1640) |
|-----|----------------------|----------------------------|----------------------------|
| 1:2 | Average Recovery (%) | 99.0 | 98.9 |
| | Range (%) | 94.4-107.6 | 92.5-105.3 |
| 1:4 | Average Recovery (%) | 96.5 | 95.7 |
| | Range (%) | 93.4-103.4 | 89.6-102.6 |

| | | | |
|------|----------------------|-----------|------------|
| 1:8 | Average Recovery (%) | 92.5 | 92.0 |
| | Range (%) | 83.0-96.5 | 87.5-100.3 |
| 1:16 | Average Recovery (%) | 89.0 | 91.4 |
| | Range (%) | 84.4-93.0 | 89.1-93.5 |

Note: The example data is for reference only.

PLATE LAYOUT



Note: Blank is a Blank Dilution Buffer hole.

TROUBLESHOOTING GUIDE

| Problem | Cause | Solution |
|---------------------|----------------------------------|--------------------------------------|
| Poor standard curve | * Inaccurate pipetting | * Check pipettes |
| Large CV | * Inaccurate pipetting | * Check pipettes |
| | * Air bubbles in wells | * Remove bubbles in wells |
| High background | * Plate is insufficiently washed | * Review the manual for proper wash. |
| | * Contaminated wash buffer | * Make fresh wash buffer |

| | | |
|---|--|--|
| <p>Very low readings across the plate</p> | <ul style="list-style-type: none"> * Incorrect wavelengths * Insufficient development time | <ul style="list-style-type: none"> * Check filters/reader * Increase development time |
| <p>Samples are reading too high, but standard curve looks fine</p> | <ul style="list-style-type: none"> * Samples contain cytokine levels above assay range | <ul style="list-style-type: none"> * Dilute samples and run again |
| <p>Drift</p> | <ul style="list-style-type: none"> * Interrupted assay set-up * Reagents not at room temperature | <ul style="list-style-type: none"> * Assay set-up should be continuous - have all standards and samples prepared appropriately before commencement of the assay * Ensure that all reagents are at room temperature before pipetting into the wells unless otherwise instructed in the antibody inserts |