

## **Human Th1/Th2 Cytokine Kit**

### **(Flow Cytometry Multiplex Bead Assay)**

#### **【Product Name】**

Human Th1/Th2 Cytokine Kit (Flow Cytometry Multiplex Bead Assay)

#### **【Catalog】**

FCM-C05R

#### **【Size】**

96 Tests

#### **【Background】**

Cytokines play an important role in physiological and pathological processes such as immune regulation, inflammatory response and tumor metastasis. Cytokine detection was widely used in infectious diseases, autoimmune diseases, tumor auxiliary diagnosis, disease evaluation, medication guidance and prognosis judgment. Multiple cytokines can be detected by the flow cytometry multiplex bead assay at the same time, and multiple results can be obtained in a single sample test at once time, reducing samples and reagents using. The compatible flow cytometers are universal and simple to operate.

#### **【Principle】**

Human Th1/Th2 Cytokine Kit (Flow Cytometry Multiplex Beads Assay) (Catalog No. FCM-C05R) gives quantitative results of Interleukin-2 (IL-2), Interleukin-4 (IL-4), Interleukin-6 (IL-6), Interleukin-10 (IL-10), Tumor Necrosis Factor  $\alpha$  type (TNF- $\alpha$ ) and Interferon- $\gamma$  (IFN- $\gamma$ ) with a single sample testing. The performance of this kit has been optimized for specific analysis of cytokines in cell culture supernatants, plasma and serum samples. The kit provides sufficient reagents for the quantitative analysis of 96 tests.

The antibody encapsulated 6-plex beads, protein (IL-2/4/6/10, TNF- $\alpha$ , IFN- $\gamma$ ) in specimens and detection antibody formed a sandwich complex as bead-analyte-detection antibody. The intensity of PE fluorescence, in proportion to the recombinant protein titer in specimens, was assessed by flow cytometry at wavelength of 575 nm approximately. The intensity of APC fluorescence was applied to classify bead populations, at wavelength of 670 nm approximately.

## 【Components】

Table 1. Materials and Storage

| ID         | Components                        | Size (96 tests)                 | Format           | Storage (Unopened)            | Storage (Opened)              |
|------------|-----------------------------------|---------------------------------|------------------|-------------------------------|-------------------------------|
| FCM05-C01  | Calibrator                        | 5 $\mu$ g (for each) $\times$ 2 | Powder           | 2-8 $^{\circ}$ C              | -20 $^{\circ}$ C              |
| FCM05-C02A | Human IL-2 Capture Beads          | 96 tests                        | Beads suspension | 2-8 $^{\circ}$ C, avoid light | 2-8 $^{\circ}$ C, avoid light |
| FCM05-C02B | Human IL-4 Capture Beads          | 96 tests                        | Beads suspension | 2-8 $^{\circ}$ C, avoid light | 2-8 $^{\circ}$ C, avoid light |
| FCM05-C02C | Human IL-6 Capture Beads          | 96 tests                        | Beads suspension | 2-8 $^{\circ}$ C, avoid light | 2-8 $^{\circ}$ C, avoid light |
| FCM05-C02D | Human IL-10 Capture Beads         | 96 tests                        | Beads suspension | 2-8 $^{\circ}$ C, avoid light | 2-8 $^{\circ}$ C, avoid light |
| FCM05-C02E | Human TNF- $\alpha$ Capture Beads | 96 tests                        | Beads suspension | 2-8 $^{\circ}$ C, avoid light | 2-8 $^{\circ}$ C, avoid light |
| FCM05-C02F | Human IFN- $\gamma$ Capture Beads | 96 tests                        | Beads suspension | 2-8 $^{\circ}$ C, avoid light | 2-8 $^{\circ}$ C, avoid light |
| FCM05-C03  | 2 $\times$ Assay Buffer           | 40 mL                           | Liquid           | 2-8 $^{\circ}$ C              | 2-8 $^{\circ}$ C              |
| FCM05-C04  | 10 $\times$ Wash Buffer           | 10 mL                           | Liquid           | 2-8 $^{\circ}$ C              | 2-8 $^{\circ}$ C              |
| FCM05-C05  | Detection Antibody                | 0.5 mL                          | Liquid           | 2-8 $^{\circ}$ C, avoid light | 2-8 $^{\circ}$ C, avoid light |
| FCM05-C06  | 96-well V-bottom Plate            | 1 plate                         | Solid            | 2-8 $^{\circ}$ C              | 2-8 $^{\circ}$ C              |
| FCM05-C07  | 96-well Sealing Film              | 2 pieces                        | Solid            | 2-8 $^{\circ}$ C              | 2-8 $^{\circ}$ C              |
| FCM05-C08  | APC Positive Control              | 0.5 mL                          | Beads suspension | 2-8 $^{\circ}$ C, avoid light | 2-8 $^{\circ}$ C, avoid light |
| FCM05-C09  | PE Positive Control               | 0.5 mL                          | Beads suspension | 2-8 $^{\circ}$ C, avoid light | 2-8 $^{\circ}$ C, avoid light |

## 【Storage and Expiration】

1. Unopened kit should be stored at 2 °C~8 °C upon receiving.
2. The expiration date is labeled on the package box. DO NOT use reagents beyond expiration date.
3. The opened kit should be stored per component, as indicated in Table 1. The shelf life of all components and dilution components are 30 days from the date of opening.

*Note: Freeze and thaw NO MORE THAN 2 times, once calibrator (ID# FCM05-C01) is reconstituted.*

## 【Unsupplied Materials and Instruments】

1. Single-channel pipettes, multi-channel pipettes and pipette tips
2. Reagent reservoirs for multichannel pipette
3. Polypropylene microcentrifuge tubes for samples collection or dilution
4. Deionized or distilled ultrapure water
5. 96-well magnetic separation rack (Magnetic Capture Plate, catalog FCM-C03M)
6. Horizontal orbital shaker for 96-well plate
7. Vortex mixer
8. Flow cytometer equipped with two lasers:
  - (1) Excitation at 488 nm or 532 nm, emission around 575 nm;
  - (2) Excitation around 633 nm, emission around 670 nm

## 【Important】

1. For research use only, not for use in diagnostic or therapeutic procedures.
2. Please follow the instructions strictly, for optimal and consistent data output.
3. Protect beads suspension, detection antibody from light all times to prevent photobleaching.
4. DO NOT mix or substitute reagents from different kit lots. DO NOT mix up or substitute reagents from different manufacturers.
5. Bring the kit components to room temperature before use. Be sure the crystal precipitates are all dissolved before use.
6. Prepare the buffer, reagents, calibrator, samples and all relevance, just prior to use.

7. Deionized or distilled water must be used for reagent preparation.
8. Ensure reagent reservoirs are clean.
9. In order to avoid microbial contamination or cross-contamination of reagents or specimens which may invalidate the test, use disposable plastic pipette tips.
10. Avoid long-term storage and repeated freeze-thaw cycles of reconstituted calibrator.

## **【Precaution】**

All chemicals should be considered as potentially hazardous. It is recommended that this kit is handled only by those persons who have been trained in laboratory techniques and it is used in accordance with the principles of good laboratory practice. Suitable protective clothing such as laboratory overalls, safety glasses and gloves is needed. Attention should be paid to avoid contact with skin or eyes. In the case of contact with skin or eyes, wash immediately with plenty of water. All blood components and biological materials should be handled and disposed properly, in accordance with local and national guideline.

## 【Procedure】

### 1. Assay Preparation

#### 1.1 Preparation of Samples

##### 1.1.1 Cell culture supernatant

Fresh collected or -80 °C stored medium 4000 g centrifuge for 10 minutes at 4 °C, aspirate the supernatant and used for the next assay.

##### 1.1.2 Serum collection and storage

**1.1.2.1** Fresh blood samples were obtained from venous, keep at room temperature for more than 30 minutes. After coagulation, 2, 000 g centrifuge for 10 minutes at 4 °C (Excessive centrifugation might lead to hemolysis). Aspirate the serum layer and avoid the contamination of blood cells.

**1.1.2.2** Serum layer was centrifuged 16, 000 g for 10 minutes at 4 °C. Discard the precipitates and the supernatant was the serum freshly prepared.

**1.1.2.3** Use the serum immediately or keep at -80 °C for long time storage.

##### 1.1.3 Plasma collection and storage

**1.1.3.1** Fresh blood samples were obtained from venous, adding anticoagulant sodium citrate, such as EDTA or heparin. 2, 000 g centrifuge for 10 minutes at 4 °C. Carefully aspirate the plasma layer, and avoid the contamination of blood cells.

**1.1.3.2** Centrifuge the plasma layer 16, 000 g for 10 minutes at 4 °C. Discard the precipitates, and keep supernatant as the freshly prepared plasma. Use the plasma immediately or storage at -80 °C.

**Note 1:** Frozen serum, plasma or medium should be mixed thoroughly after thawing, and remove all visible debris by centrifuge. Thawed samples should be used up and avoid repeated freeze-thaw cycles.

**Note 2:** Hemolyzed, icteric and lipemic samples are not validated for use in this assay.

## 1.2 Preparation of Reagents and Buffer

For a repeatable detection assay, we recommend bring the kit to room temperature and keep 15 minutes before use for a temperature balancing.

### 1.2.1 Assay Buffer working solution

Aspirate 40 mL 2 × Assay Buffer (ID# FCM05-C03), mixed with 40 mL deionized water.

*Note: Assay Buffer is used for preparation of calibrator, samples and detection antibody.*

### 1.2.2 Detection Antibody working solution

Aspirate 0.5 mL Detection antibody (ID# FCM05-C05), mixed with 10.5 mL Assay Buffer.

*Note: The preparation of detection can be regulated according to samples and a freshly prepared detection antibody is recommend for a better performance.*

### 1.2.3 Wash Buffer working solution

Aspirate 10 mL 10 × Wash Buffer (ID# FCM05-C04), mixed with 90 mL deionized water.

*Note: Wash Buffer is used for reaction product washing and beads resuspended.*

### 1.2.4 Beads suspension working solution

Define cytokine to be analyzed, and fetch corresponding cytokine beads from the kit (from ID# FCM05-C02A to ID# FCM05-C02F). Vortex beads suspension vigorously no less than 30 seconds. Immediately transfer required volume of beads to a microcentrifuge tube, and mix with required volume of Assay Buffer.

*Note: DO NOT aspirate beads suspension less than 20 µL to minimize pipetting errors. Perform a serial dilution if only a few beads needed for the assay.*

We recommend a freshly prepared reagents for usage of the beads, and to setup ONE test in ONE well in 96-well V-bottom plate, add beads suspension for EACH plex, then replenish with Assay Buffer to total volume of 120 µL, as indicated in Table 2.

**Table 2. Preparation of Beads Suspension Working Solution for One Test**

| Multiplex | Single Capture Coating Beads, ID# FCM05-C02* | Add Assay Buffer to |
|-----------|--|---------------------|
| 6-plex    | 1 µL +1 µL + 1 µL+ 1 µL+ 1 µL+ 1 µL          | 120 µL              |

## 1.3 Preparation of Calibrator

**1.3.1** Reconstitute lyophilized calibrator powder with 500 µL deionized water, as calibrator stock#1 with each analyte 10 µg/mL respectively. For completely dissolving, keep the bottle at room temperature at least for 15 minutes.

*Note: Mix or reconstitute protein reagent gently, avoid bubbles and foam.*

**1.3.2** For multiplex assay, add in 900  $\mu$ L Assay Buffer in a new tube, aspirate 100 uL calibrator stock#1, labeled as calibrator stock#2. The concentration of each analyte is 1  $\mu$ g/mL respectively.

**1.3.3** Repeat operation of step 1.3.2 and labeled as stock#3 and Cal 11 respectively.

**1.3.4** Performing 2-fold serial dilutions from Cal 11, add 500  $\mu$ L Assay Buffer, labeled as Cal 10, Cal 9, Cal 8, Cal 7, Cal 6, Cal 5, Cal 4, Cal 3, Cal 2 and Cal 1 respectively, as shown in Table 3.

*Note: Mix thoroughly before making the next dilution.*

**Table 3. Preparation of Calibrator**

| Calibrator ID | Serial Dilution | Assay Buffer add in ( $\mu$ L) | Calibrator add in ( $\mu$ L) | Final Concentration (pg/mL) |
|---------------|-----------------|--------------------------------|------------------------------|-----------------------------|
| Stock#2       | 10              | 900                            | 100 $\mu$ L of stock#1       | 1,000,000                   |
| Stock#3       | 10              | 900                            | 100 $\mu$ L of stock#2       | 100,000                     |
| Cal 11        | 10              | 900                            | 100 $\mu$ L of stock#3       | 10,000                      |
| Cal 10        | 2               | 500                            | 500 $\mu$ L of Cal 11        | 5,000                       |
| Cal 9         | 2               | 500                            | 500 $\mu$ L of Cal 10        | 2,500                       |
| Cal 8         | 2               | 500                            | 500 $\mu$ L of Cal 9         | 1250                        |
| Cal 7         | 2               | 500                            | 500 $\mu$ L of Cal 8         | 625                         |
| Cal 6         | 2               | 500                            | 500 $\mu$ L of Cal 7         | 312.5                       |
| Cal 5         | 2               | 500                            | 500 $\mu$ L of Cal 6         | 156.3                       |
| Cal 4         | 2               | 500                            | 500 $\mu$ L of Cal 5         | 78.1                        |
| Cal 3         | 2               | 500                            | 500 $\mu$ L of Cal 4         | 39.1                        |
| Cal 2         | 2               | 500                            | 500 $\mu$ L of Cal 3         | 19.5                        |
| Cal 1         | 2               | 500                            | 500 $\mu$ L of Cal 2         | 9.8                         |
| Cal 0         | -               | 500                            | -                            | 0                           |

## 2. Assay Procedure

**2.1** Plasma/serum preparation (one sample repeat). Dilute 30  $\mu\text{L}$  freshly prepared plasma or thawed serum with 60  $\mu\text{L}$  Assay Buffer, mix homogenously and ready for being used.

*Note1: Plasma/serum samples dilution may need to be regulated according to the concentration of the cytokines.*

*Note2: Medium matrix affection has been investigated and freshly prepared medium can be detected directly, for a better accuracy detection we recommend a suitable regulated dilution.*

**2.2** Add serial dilutions of calibrator or samples to 96-well V-bottom plate, 30  $\mu\text{L}$  per well.

*Note: Run calibrators in duplicates. Follow the attached plate layout to achieve good accuracy.*

**2.3** Add beads suspension working solution to 96-well V-bottom plate, 120  $\mu\text{L}$  per well.

**2.4** Seal the plate. Incubate at room temperature for 120 minutes, with continuous shaking 400-600 rpm to ensure the beads always suspended homogenously in the solution. Avoid light.

**2.5** Place the 96-well V-bottom plate onto the magnetic separation rack for 2 minutes before discarding supernatant and remove plate from separation rack.

*Note: Magnet varies in strength. It may take a few seconds to minutes, to complete the separation.*

**2.6** Reconstitute each well in 200  $\mu\text{L}$  of Wash Buffer. Mix thoroughly using pipette by aspirating and dispensing 2-3 times. Place the 96-well V-bottom plate onto the magnetic separation rack for 2 minutes before discarding supernatant and remove the plate from the magnet.

**2.7** Add detection antibody, 100  $\mu\text{L}$  per well.

**2.8** Seal the plate. Incubate at room temperature for 60 minutes, with continuous shaking 400-600 rpm to ensure the beads always suspended homogenously in the solution. Avoid light.

**2.9** Place the 96-well V-bottom plate onto the magnetic separation rack for 2 minutes before discarding supernatant and remove plate from separation rack.

**2.10** Repeat step 2.6

*Note: For a low blank signal, the washing can be repeated one more time.*

**2.11** Add 150  $\mu\text{L}$  Wash Buffer to each well. Mix by pipetting up and down. Ensure the beads well



separated and not aggregated.

**2.12** Subject to flow cytometry analysis. If not being analyzed immediately, store at 2~8 °C and avoid light. Flow cytometry assay should be performed within 2 hours.

*Note: Resuspend beads immediately prior to reading by pipetting up and down.*

### 3. Flow Cytometer Setup

**3.1** Flow cytometer equipped with two lasers are compatible with the assay

(1) excitation laser at 488 nm or 532 nm, and emission around 575 nm;

(2) excitation laser around 633 nm, and emission around 670 nm.

Instruments tested by this assay were represented in Table 4.

**Table 4. Partial List of Compatible Flow Cytometers**

| Manufacturer             | Verified instrument model | Classification Channel | Reporter Channel |
|--------------------------|---------------------------|------------------------|------------------|
| BD Biosciences           | BD FACSLyric™             | APC                    | PE               |
| Beckman Coulter          | Cytoflex S                | R660-APC               | Y585-PE          |
| Thermo Fisher Scientific | Attune NxT                | RL1                    | YL1              |
| Luminex Corporation      | Guava easy Cyte3L         | RED-R                  | YEL-B            |

**3.2** Channel setup

**3.2.1** PE Positive Control vortex 30 sec, subject to flow cytometry. For the voltage setup of the reporter channel, not as medium flow rate in samples running, we recommend a low rate, and 8E5 (Beckman Coulter, Cytoflex S) as a threshold value for the PE signal.

**3.2.2** APC Positive Control using for the setup of the classification channel as the PE Beads, the APC signal located at right range side of the detection platforms but not with an outside distribution is an optimal situation.

**3.3** Select medium flow rate.

**3.4** Set up 500 events or beads per plex (3,000 events for 6 plex) collected in P1 gate as stop criteria.

## 4. Data Acquisition and Analysis

### 4.1 Data acquisition

**4.1.1** Make sure the flow cytometer is well tuned, following the instrument user guide and method configuration illustrated above.

**4.1.2** Create an experiment in 96-well plate format.

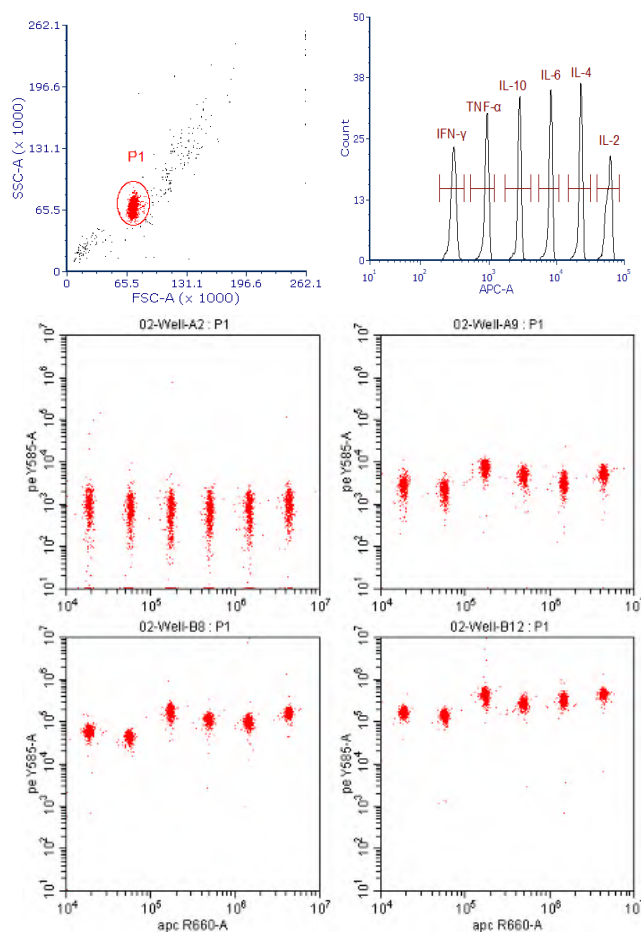
*Note: If 96-well plate loader is not available, transfer the samples to FACS tubes, replenish with 100  $\mu$ L Wash Buffer, and read one by one.*

**4.1.3** Resuspend beads by pipetting up and down.

**4.1.4** Load the plate and start acquisition.

**4.1.5** In APC-count histogram, create P2 ~ P7 gates in P1 sub-population. Six-plex beads was differentiated, as indicated in Table 5.

**Table 5. Standards of Analyte-specific Populations**



**4.1.6** Record median fluorescence intensity (MFI) of PE channel.

**4.2** Data analysis

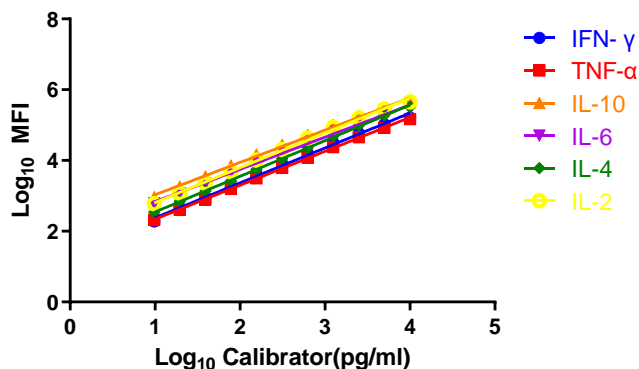
**4.2.1** The following data, acquired by BECKMAN Cytoflex, as indicated in Table 5.

**4.2.2** Two-log-linear fit curve model is applied by data analysis with GraphPad by plotting Log<sub>10</sub> concentration value of serial diluted calibrators against Log<sub>10</sub> median fluorescence intensity (MFI) of PE channel, or you can use the FCAP for data analysis. We recommend the r<sup>2</sup> value of the curve above 0.99.

*Note: If two-log-linear fit curve model is not suitable for the data analysis, other regression such as four parameters can be another choice.*

**4.2.3** Standard curve examples, the following graphs represent standard curves from the Human Th1/Th2 Cytokine Kit (Flow Cytometry Multiplex Bead Assay) (Catalog No. FCM-C05R), as indicated in Table 6.

**Table 6. Standard Curves**



**4.2.4** Calculate the concentration of unknown from the calibration curve of each analyte.

**【Quick Guide】**



**1** 96-well plate  
 ★ Add 30  $\mu$ L calibrator or samples  
 ★ Add 120  $\mu$ L beads suspension working solution

Shake, avoid light 18-25  $^{\circ}$ C 120 minutes

Place the 96 well plate on magnetic separation rack and stay for 2 minutes. Discard supernatant.



**2** ★ Wash with 200  $\mu$ L Wash Buffer, pipetting up and down for 2-3 times



Place the 96 well plate on magnetic separation rack and stay for 2 minutes. Discard supernatant.



**3** ★ Add 100  $\mu$ L detection antibody working solution

Shake, avoid light 18-25  $^{\circ}$ C 60 minutes

Place the 96 well plate on magnetic separation rack and stay for 2 minutes. Discard supernatant.



**4** ★ Repeat Wash step.  
 ★ Place the 96 well plate on magnetic separation rack and stay for 2 minutes. Discard supernatant.  
 ★ Add 150  $\mu$ L Wash Buffer to each well. Mix by pipetting up and down.



**5** ★ Flow cytometry analysis. If not being analyzed immediately, store at 2~8  $^{\circ}$ C, avoid light and should be performed within 2 hours.

## 【Trouble Shooting】

| Concerns  | Possibilities  | Suggestions   |
|---|--|---|
| After magnetic separation, the magnetic beads precipitates are not visible or become less and less after multiple-step operation. | The pellets are very loosely attached to the well, and lost during aspiration.           | Aspirate the supernatant slowly and carefully. Keep the plate at magnet for a long time with a visible accumulation of the beads.     |
| Variation of beads count in duplicated wells.   | Aspiration takes so long time that the beads settled to the bottom of the tube or wells. | Quickly aspirate and dispense the bead suspension. Vortex beads vigorously before first use, and vortex briefly in between operation. |
| Plenty of debris were observed in FSC-SSC scatter plot during data acquisition.   | Improper setting of FSC and SSC threshold.   | Increase threshold value of FSC and SSC.  |
| Plenty of beads doublets are observed by plotting FSC height versus FCS area.   | Beads aggregate due to long time sitting or insufficient resuspending.                   | Resuspend the beads by pipetting up and down vigorously, then re-load onto flow cytometer.  |
| Less than 6 bead populations in APC-count histogram.  | The PMT gain or voltage value of APC fluorescent channel is too high.                    | Adjust PMT gain or voltage of APC fluorescent channel, ensuring 6 intact peaks observed.  |
| Less than 6 bead populations in APC-PE scatter plot, though all the 6 peaks obtained in APC-count histogram.                      | The PMT gain or voltage value of PE fluorescent channel is too high.                     | Adjust PMT gain or voltage of PE fluorescent channel, ensuring 6 bead populations in APC-PE scatter plot.                             |
| PE fluorescent intensity of low concentration calibrator is higher than that of high concentration calibrator.                    | Insufficient needle wash and clean between samples.                                      | At least one washing cycle between samples in flow cytometer setting.   |
|   |  | Follow the Plate Layout suggested, and read the plate by columns to reduce cross-contamination on flow cytometer.                     |

## 【Plate Layout】

Arrange the samples in vertical, and read the plate by column.

|   | 1     | 2     | 3      | 4        | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
|---|-------|-------|--------|----------|---|---|---|---|---|----|----|----|
| A | Blank | Cal 4 | Cal 8  | Sample 1 |   |   |   |   |   |    |    |    |
| B | Blank | Cal 4 | Cal 8  | Sample 1 |   |   |   |   |   |    |    |    |
| C | Cal 1 | Cal 5 | Cal 9  | Sample 2 |   |   |   |   |   |    |    |    |
| D | Cal 1 | Cal 5 | Cal 9  | Sample 2 |   |   |   |   |   |    |    |    |
| E | Cal 2 | Cal 6 | Cal 10 | .....    |   |   |   |   |   |    |    |    |
| F | Cal 2 | Cal 6 | Cal 10 | .....    |   |   |   |   |   |    |    |    |
| G | Cal 3 | Cal 7 | Cal 11 | .....    |   |   |   |   |   |    |    |    |
| H | Cal 3 | Cal 7 | Cal 11 | .....    |   |   |   |   |   |    |    |    |